



RESEARCH ARTICLE

A STUDY ON CORD BLOOD BILIRUBIN LEVELS IN A TERTIARY CARE CENTRE OF HARYANA IN INDIAGurdeep Singh Dhanjal¹, Latika Sahni², P.D. Sharma³¹ Assistant Professor, Department of Pediatrics, Maharishi Markandeshwar University, Mullana, Distt. Ambala (Haryana) India.² P.G. - 3rd Year, Department of Pediatrics, Maharishi Markandeshwar University, Mullana, Distt. Ambala (Haryana) India.³ Professor and Head, Department of pediatrics, Maharishi Markandeshwar University, Mullana, Distt. Ambala (Haryana), India.

Received 05 September 2013; Revised 20 September 2013; Accepted 22 September 2013

ABSTRACT

To determine the level of cord blood bilirubin in all healthy term newborns and assess its usefulness in predicting neonatal jaundice. Neonatal jaundice (NNJ) is an interesting, complicated and controversial clinical problem. It is a cause of concern for the parents as well as pediatricians. The concept of prediction of jaundice offers an attractive option to pick up babies at risk of neonatal hyperbilirubinemia. Physical examination is not a reliable measure of serum bilirubin. Under these circumstances it would be desirable to be able to predict the risk of jaundice, in order to implement early treatment and thereby minimize the risk of bilirubin dependent brain damage. Neonatal Hyperbilirubinemia has been defined as the bilirubin levels > 12.9 mg/dl in term babies and 15 mg/dl in preterm babies. Neonatal jaundice is visible manifestation in skin and sclera of elevated serum concentrations of bilirubin and this usually occurs in neonates if serum bilirubin level is >5 mg/dl. Most adults are jaundiced when total serum bilirubin (TSB) levels exceed 2.0 mg/dL. Kernicterus and near miss kernicterus are neonatal conditions that are associated with irreversible or reversible brain injury respectively.⁵ Concerns regarding jaundice have increased after reports of bilirubin encephalopathy occurring in healthy term infants without hemolysis. The study was conducted in 100 term newborns in neonatal unit, Department of Pediatrics, Maharishi Markandeshwar Institute of Medical Sciences and Research (MMIMSR), Mullana, from September 2012- July 2013. Serum bilirubin estimation was done in the biochemistry department of MMIMSR, Mullana. Serum bilirubin estimation was done by Diazo method in which a detergent is used to accelerate the reaction is used. Incidence of NNJ in our study is 14 %. Mean total bilirubin on third post natal day was 9.14 mg/dl. Using CBB level of ≥ 1.9 mg/dl as a cut-off, NNJ can be predicted with sensitivity of 92.8%, specificity of 83.7 %, positive predictive value of 48.1 % and negative predictive value of 98.6 %. The Negative Predictive Value (98.6%) in the present study suggests that in healthy term babies (without RH and ABO incompatibility with Cord Blood Bilirubin ≤ 1.9 mg/dl) cord serum bilirubin can help to identify those newborns who are unlikely to require further evaluation and intervention. These newborns can be discharged with assurance to parents. Babies with CBB level ≥ 1.9 mg/dl should be followed more frequently.

Keywords: Newborn, Neonate, Neonatal Jaundice, Cord blood bilirubin.**INTRODUCTION:**

Health is defined as a state of complete physical, mental and social well being and not merely an absence of disease. Health is a fundamental human right¹. In spite of completing more than 60 years of independence and with so many prevailing health programmes, Infant Mortality rate is high compared to developed countries. Infant mortality rate is one of the most universally accepted indicator of health status not only of infants but also the whole population².

Developing countries like India must be fully aware of this limitation on the development of neonatal care, particularly neonatal intensive care. The ultimate aim

should be to benefit maximum number of newborn babies with improved survival and reduced mortality.

Clinical jaundice is seen in 60-70% of term and about 80% of preterm newborns³. 6.1% of well term newborn have a serum bilirubin over 12.9mg%. Serum bilirubin over 15 mg% is found in 3 % of normal term newborns.⁴ The potential risk of developing bilirubin encephalopathy or even kernicterus is high in babies with elevated serum bilirubin level. The sequelae could be serious as patients may develop cerebral palsy, sensorineural deafness and mental retardation⁵. Concerns regarding jaundice have increased after reports of bilirubin encephalopathy occurring in healthy term infants without hemolysis⁶.

Factors like a why worry attitude about jaundice among doctors, financial constraints, family and medical consideration, shortage of hospital beds and personnel frequently influence the decision about early hospital discharge of the mother and infant after birth⁷. Total serum bilirubin (TSB) in infants discharged within 48 hours of life, generally shows an increasing trend and some of these infants later develop significant hyperbilirubinemia requiring treatment⁸.

During past 100 years, few major themes have dominated the thinking about pathogenesis and treatment of neonatal jaundice beginning with an understanding of the chemical basis of jaundice and identification of the substance causing jaundice as bilirubin. Bilirubin when conventionally illustrated has ridge like conformation⁹.

Bilirubin is derived from the heme containing proteins in the reticuloendothelial system. The normal newborn produces 6 to 10 mg of bilirubin per kilogram per day that is nearly 2 to 3 times as compared to adult. The destruction of circulating red blood cells accounts for 75% of daily bilirubin production in the neonate¹⁰. 1 gm of hemoglobin produces 34mg of bilirubin. The remaining 25% of bilirubin called early labeled bilirubin¹¹. It is derived from ineffective erythropoiesis in bone marrow from other heme containing proteins in tissues (e.g., myoglobin, cytochromes, catalases and peroxidases and from free heme).¹² The heme ring from heme containing proteins is oxidized to bilirubin by heme oxygenase. This enzyme is called the *rate limiting enzyme*. Heme oxygenase is a microsomal enzyme found in reticuloendothelial system and also in tissue macrophages and in gut mucosa, liver, spleen. In the fetus, hepatic microsomal heme oxygenase activity has been measured to be 8 times higher than in adult liver. Heme oxygenase must be present by the time bilirubin appears in human fetus at 14 weeks gestation. The subsequent step, reduction at the C-11 carbon of biliverdin IX α to bilirubin IX α takes place in cytosol and it is catalyzed by biliverdin reductase¹³. Both of these processes are dependent on nicotinamide adenine dinucleotide phosphate oxidase (NADPH). This reaction releases carbon monoxide (CO) (excreted from lung) and iron (reutilized). Catabolism of 1 mmol of hemoglobin produces 1 mmol each of CO and bilirubin. The metabolism of heme results in the formation of free iron, CO and biliverdin, which in turn is reduced to bilirubin¹⁴. Unconjugated bilirubin is transported in plasma bound to albumin with a binding affinity of 10^7 to $10^8 M^{-1}$ at the primary binding site. This bound form does not cross the blood brain barrier and is nontoxic¹⁵. In addition to albumin, bilirubin can also bind to other proteins (e.g.,

alpha-fetoprotein and ligandin) as well as to lipoproteins. Lysine is involved in bilirubin binding to both albumin and ligandin and possibly to other proteins. Recent findings suggest that binding to lysine may play a role in mediation or modulation of bilirubin toxicity¹⁶.

Bilirubin dissociates from albumin at the sinusoidal surface of hepatocyte, being taken up by facilitated diffusion. Inside the hepatocyte bilirubin binds to cytosolic *glutathione-s-transferases* initially termed ligandins. Binding to glutathione-s-transferases keeps unconjugated bilirubin soluble in cytosol of hepatocyte and increases the net uptake of bilirubin by reducing its efflux from the cell¹⁷. Conjugation (glucuronidation) of bilirubin is essential for the efficient biliary excretion of bilirubin. Bilirubin glucuronidation is catalyzed by specific isoform of *uridinediphosphoglucuronate glucuronyl transferase*, termed UGT1A1. It catalyzes the binding first one molecule of glucuronic acid to bilirubin, forming bilirubin monoglucuronide. Conjugated bilirubin is not absorbed from the intestine, but the small amount of unconjugated bilirubin that appears in the bile is partially reabsorbed. The gut bacteria reduce the conjugated bilirubin to stercobilinogen, which is excreted with feces. The newborn is born with sterile gut and intestinal microbial flora, and there is an enzyme, called β -*glucuronidase*, which converts bile glucuronide into unconjugated bilirubin that is reabsorbed into the circulation. This is called *enterohepatic circulation* and is particularly important in babies who are not fed by mouth from birth, as with introduction of feeds bacteria destroys this enzyme¹⁸. Meconium also contains a significant amount of bilirubin estimated to be as much as 5-10 times daily production. Half of this is unconjugated and thus capable of being reabsorbed. Intestinal bacteria degrades bilirubin into urobilinogen, most of which is absorbed from intestine and undergoes enterohepatic circulation¹⁹. A minor fraction is then excreted in the urine and stool. Unconjugated bilirubin in the mammalian fetus can be disposed of either by crossing the placenta into maternal circulation or by passing through fetal liver and being excreted into fetal bile. Placental membranes are essentially impermeable to polar compounds such as biliverdin and bilirubin conjugate but non-polar compounds such as unconjugated bilirubin can diffuse across²⁰. The transplacental gradient of bilirubin from fetus to mother was between 5:1 to ~10:1. In early life, bilirubin binds to fetoprotein and liver plays minor role in the excretion. Bilirubin has been found in amniotic fluid from 12th week of gestation but gradually disappears as the volume of amniotic fluid increases by 36-37 weeks. As amniotic fluid is primarily a fetal product, it is thought that bilirubin in this fluid most likely comes from the fetus

itself²¹. The fetus swallows significant amounts of amniotic fluid and it has been speculated that this ingested bilirubin may be absorbed across the intestinal mucosa. One of the reason, the body converts biliverdin, a polar, hydrophilic, excretable, non-toxic molecule to bilirubin a non polar, hydrophobic, non excretable and

toxic molecule is that biliverdin, unlike bilirubin doesn't cross the placenta easily²². This conversion protects the fetus from biliverdin overload. For same reason the fetus has a defective hepatic bilirubin conjugated mechanism but active unconjugated mechanism²³.

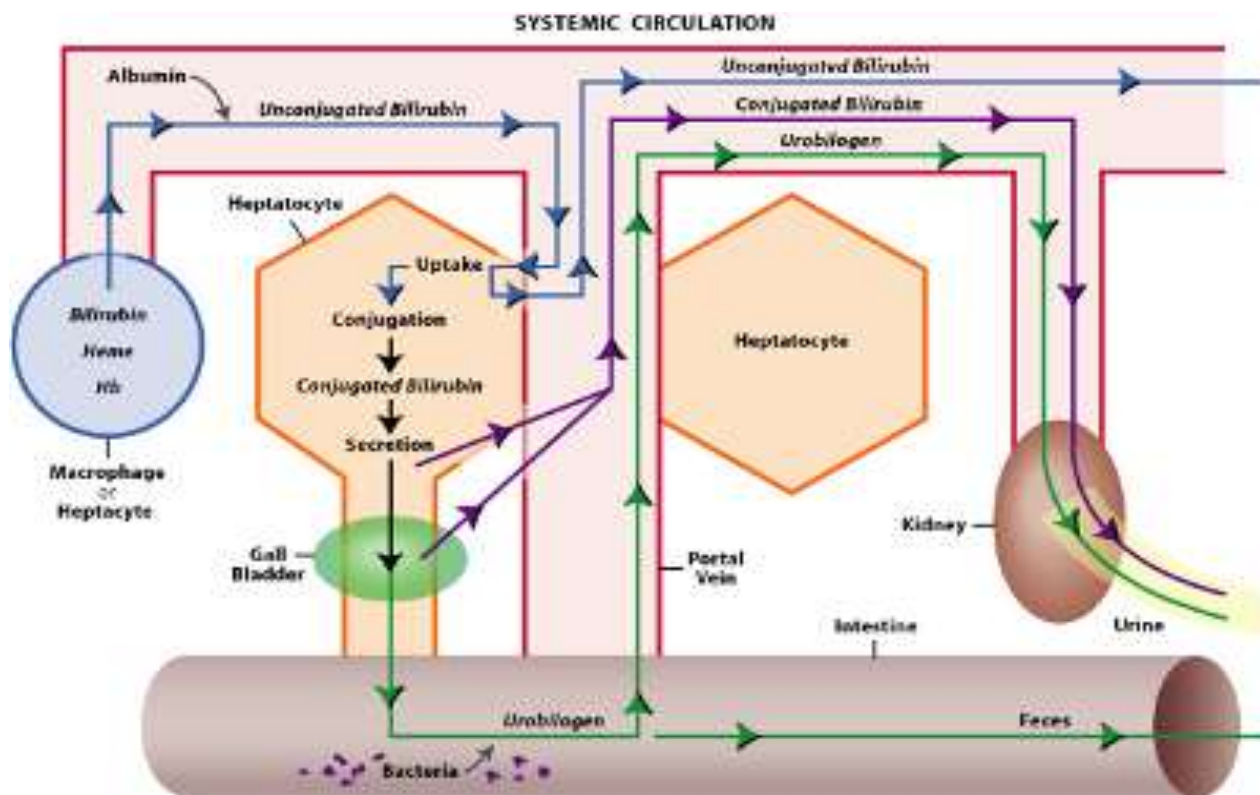


Figure 1: Schematic illustration of bilirubin metabolism in fetal and neonatal life

MATERIALS AND METHODS:

This study was conducted in 100 term newborns in Neonatal unit, Department of Pediatrics, Maharishi Markandeshwar Institute of Medical Sciences and Research (MMIMSR), Mullana, from September 2012-July 2013. Serum bilirubin estimation was done at birth from cord blood and then at 72 hours of life from peripheral venous blood sample.

A prospective cohort study model was adopted, in which the group of newborns according to the inclusion criteria were followed up clinically and by laboratory investigation during the period of their hospital stay. At the time of this study, neonates were observed for three days post-delivery period, prior to discharge.

Inclusion Criteria was all full term neonates delivered by normal vaginal delivery (NVD) or lower segment caesarean section (LSCS), with birth weight >2.5 kg and no birth asphyxia.

The study parameter included mother's interview to record clinical history in a specially designed proforma. Cord blood samples for analysis of conjugated, unconjugated and total bilirubin levels were collected from all newborns that complied with the protocol inclusion criteria.

These newborns were followed up for the 3-day period of their hospital stay. They were physically examined daily and also whenever necessary bilirubin levels were done. Values of bilirubin at third day were compared with those of cord blood bilirubin. In all cases the recommendation for phototherapy was followed according to the schedule proposed by the AAP.²² Establishment of the mother's and newborn's blood groups were also done.

2 ml each of plain cord blood samples were collected from the umbilical cord and subjected to following investigations:

- ABO Blood grouping and Rh typing.
- Total and differential serum bilirubin assessment.

- 2 ml plain venous blood was collected from the baby after 72 hours and total and differential serum bilirubin were assessed.
- Blood collected was transported to laboratory within 2 hours of collection.

Serum bilirubin estimation was done in the biochemistry department of MMIMSR, Mullana. Serum bilirubin estimation was done by Diazo method in which a detergent is used to accelerate the reaction is used. The diazo reagent used is 2, 5 – dichlorophenyl diazonium tetrafluoroborate. This method for bilirubin estimation is based on the principle that bilirubin reacts with diazotized sulphanilic acid in acidic medium to form pink coloured azobilirubin with absorbance directly proportional to the serum bilirubin concentration. Direct bilirubin, being water soluble, directly reacts in acidic

medium. However, indirect or unconjugated bilirubin is solubilized using a surfactant and then it reacts similar to direct bilirubin.

The main outcome of this study was analyzed in terms of hyperbilirubinemia requiring phototherapy serum bilirubin level of ≥ 15 mg/dL was taken as NNJ.²²

All babies were classified into four groups depending on the UCS bilirubin levels <0.9 (group-I), 1.0-1.9 (group-II), 2.0-2.9 (group-III), >3 (group-IV). Serum bilirubin estimation was done after 72 hours of postnatal life. Babies were categorized according to the need for phototherapy.

Statistical tests of significance (chi-square) were applied and the predictive values (sensitivity, specificity, PPV, NPV) were calculated using the conventional formulae.



Figure 2: Clinical estimation of neonatal jaundice

Results and Discussion:

The incidence of hyperbilirubinemia in the present study was 14 %, 58% of which are males and 42% are females.

Incidence was more in babies born through LSCS (77.77%) than in NVD (23.23%)

Table 1: Basal parameters

Study (No.of Cases)	UCS Bilirubin (mg/dl)	Incidence of Jaundice(%)
Present study (100)	0-0.9	0
	1 – 1.9	1.9
	2 – 2.9	40.9
	>3	80

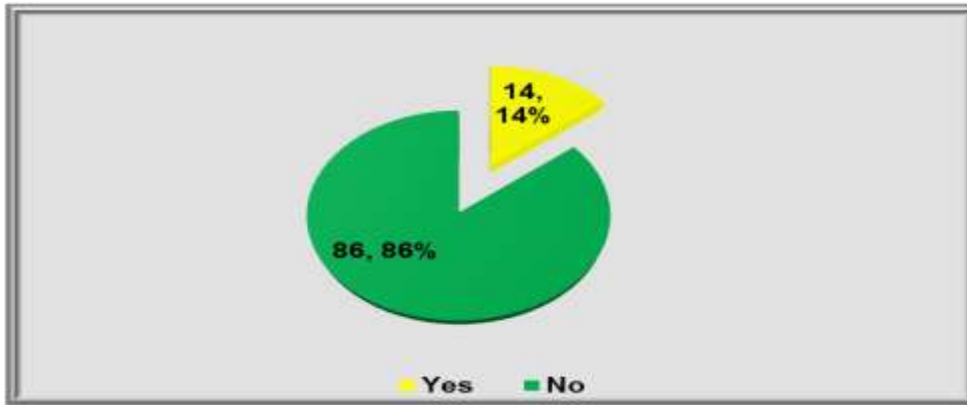


Figure 3: Distribution of study population

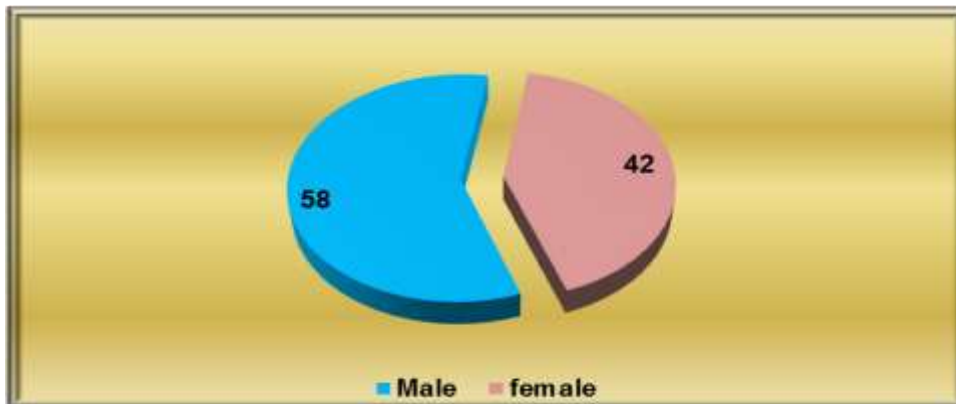


Figure 4: Sex wise distribution of study group

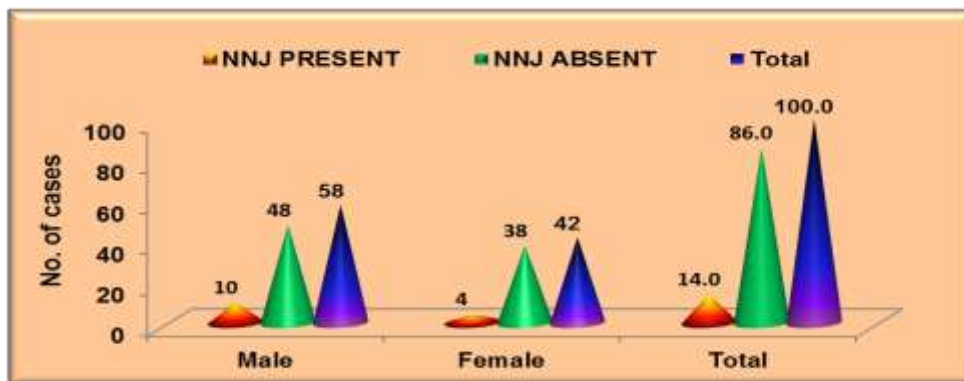


Figure 5: Association between sex of newborn and NNJ

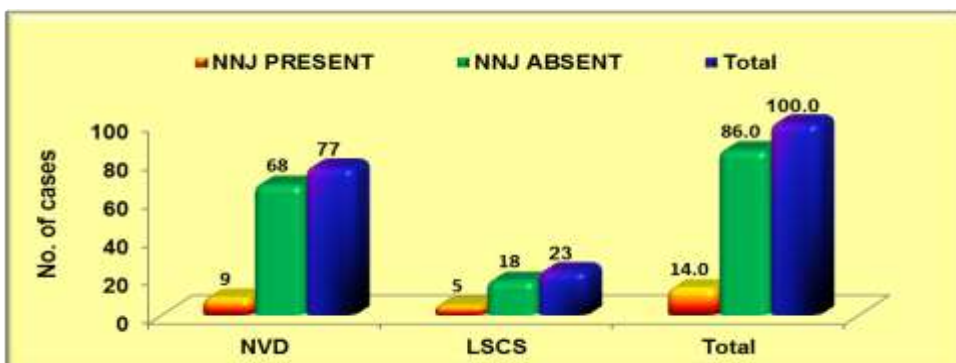


Figure 6: NNJ and Mode of Delivery

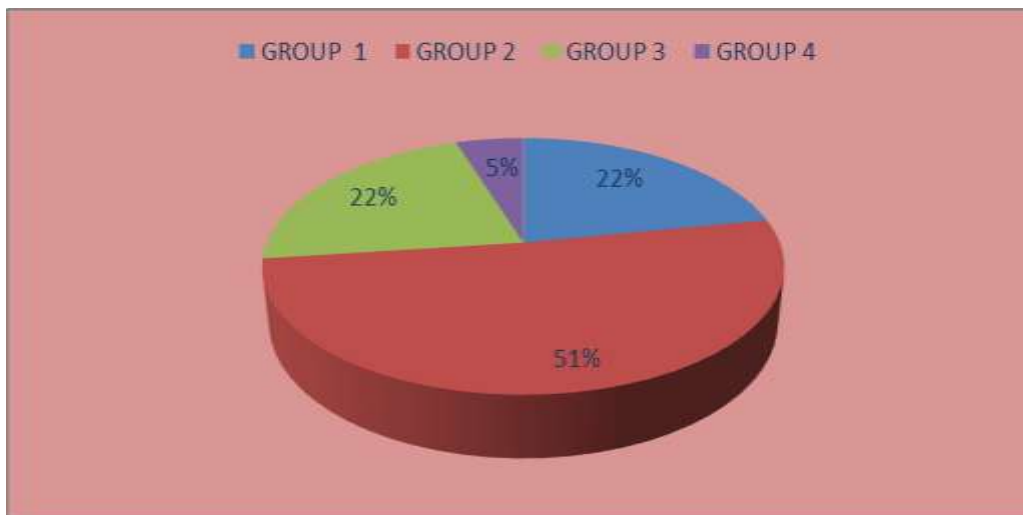


Figure 7: Incidence of NNJ in each group

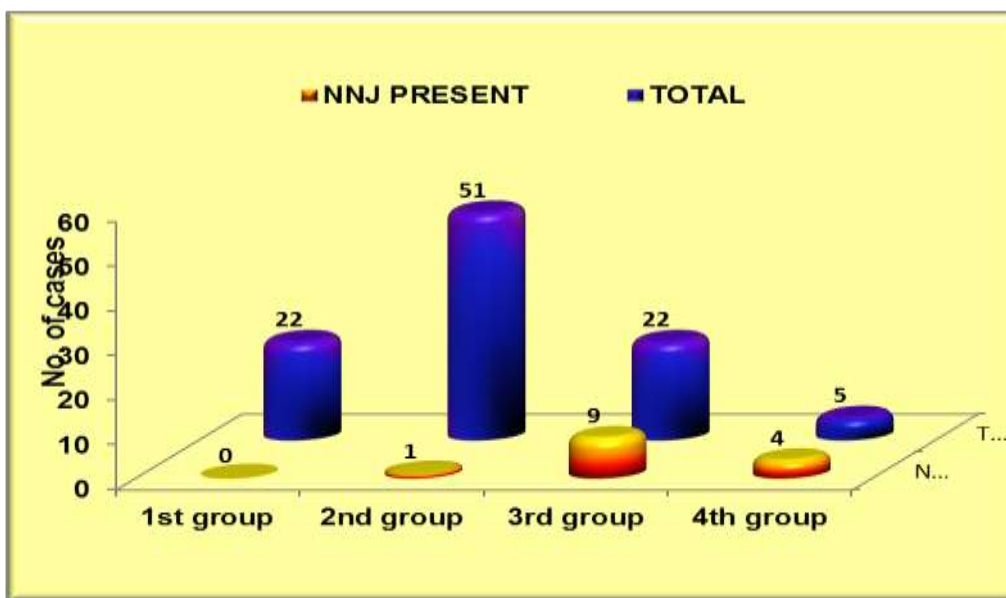


Figure 8: Incidence of NNJ in each group

Discussion and Conclusion:

The study group consisted of 100 full-terms Appropriate for gestational age neonates delivered in MMIMSR, Mullana between September 2012- September 2013. Incidence of hyperbilirubinemia was 14% in study population. Mean umbilical cord serum bilirubin was 1.63±0.73. There was no significant association between neonatal hyperbilirubinemia and route of delivery (p>0.005). No association was found between neonatal hyperbilirubinemia and sex of the baby and maternal age (p>0.05, not significant). The correlation between the mother’s blood group and the increased risk of hyperbilirubinemia in neonates was insignificant. Significant association was seen between neonatal hyperbilirubinemia and increasing umbilical cord serum bilirubin (p= 0.001).Using umbilical cord blood bilirubin

level of ≥1.9 mg/dL hyperbilirubinemia could be predicted with sensitivity of 92.8% , specificity of 83.7% , positive predictive value of 48.1% and negative predictive value of 98.6%.

Our data suggest that umbilical cord blood can be utilized for estimation of serum bilirubin to predict development of neonatal hyperbilirubinemia and decide need for appropriate intervention in healthy term neonates.

REFERENCES:

1. Concept of health and disease. K. Park. Parks textbook of preventive and social medicine 20th edtn;2001, 12-16.
2. Preventive medicine in obstetrics, paediatrics and geriatrics. K.Park. parks textbook of preventive and social medicine 20th edition 2003,12-16.

3. Anthony j piazza and Barbara J stoll. Jaundice and hyperbilirubinemia in newborn, nelson textbook of pediatrics ,18th edition 2008; 756-765.
4. Hinkes MT , cloharty jp. Neonatal hyperbilirubinemia. In Cloharty manual of neonatal care, 5th edition 1998;175-211
5. Agarwal R, Deorari A.K. Unconjugated hyperbilirubinemia in newborns: Current prospective. Indian Pediatr. 2002;39:30-42.
6. Maisels MJ, kring e ; length of stay, jaundice and hospital re admission. Pediatrics 1998; 101:995-998
7. Risemberg HM ,Mazzi E, macdonald MG, Peralta M. Heldrich F. Correlation cord bilirubin levels with hyperbilirubinemia in ABO incompatibility. Arch Dis Child 1977;57: 219-222
8. Rosenfeld J Umbilical cord bilirubin levels as predictor of subsequent hyperbilirubinemia .J Fam Pract 1986;23:556-58
9. Suchonska B. Weiglos M, Bobrowska K, Marianowski. Concentration of bilirubin in the umbilical cord blood as indicator of hyperbilirubinemia in newborns. Ginekol-Pol J 2004;75(10):749-53.
10. Mamdouha AB. The neuropathology of kernicterus: Definitions and Debate. In: Jaffrey MM and John F Watchko eds. Neonatal Jaundice. New Jersey, Harwood Academic Publishers. 2000:75-88.
11. Oski FA. The erythrocyte and its disorders. In: Nathan DG and Oski FA, Hematology of Infancy and Childhood. Philadelphia. WB Saunders Company. 2013;7:22-43.
12. Monte MJ, Rodriguez Bravo T, Macias RIR, Bravo P et al. Relationship between bile acid transport gradients and transport across the fetal facing plasma membrane of human trophoblast. Pediatr Res. 2000;38:156-63.
13. Willy T, Hansen R. Fetal and neonatal bilirubin metabolism. In: M.Jeffrey Maisles and Jon F Watchko Eds. Neonatal jaundice. Harwood Academic Publishers, New Jersey. 2000;3-20.
14. Guruprasad G. Bilirubin Metabolism- what we should know?. J Neonatol.2001;1:4-7.
15. Wolaas SI, Greengard P. Protein phosphorylation and neuronal function. Pharmacol Rev. 1999;43:299-349.
16. Hansen TWR, Mathiesen SBW, Walaus. Modulation of the effect of bilirubin on protein phosphorylation by lysine containing peptides. Pediatr Res. 1997; 42: 615-617.
17. Chowdhary JR, Wolkoff AW, Chowdhary NR, Arias IM. Hereditary jaundice disorders of bilirubin metabolism. AL Boudet, WS Shy and P Valle Eds. The metabolic and molecular basis of inherited diseases. McGraw Hill, New York; 2001;8: 3063-101.
18. Xiawang, Chowdhary JR, Chowdhary NR. Bilirubin metabolism applied physiology. Curr Pediatr, 2006;16(1): 70-4.
19. Rosenthal P. Human placental bilirubin metabolism. Pediatr Res. 1990; 27: 223A.
20. Maisles MJ. Neonatal jaundice. In: Avery GB, Hatcher MA, MacDonald MG (Eds). Neonatology, Pathology and Management of the Newborn. JB Lippincott Co. Philadelphia: 1999;5:765-820.
21. American Academy of Pediatrics Clinical Practice Guideline and Subcommittee on Hyperbilirubinemia. Management of hyperbilirubinemia in the newborn Infant 35 or More weeks of Gestation. Pediatrics. 2004 ;114:297-316.
22. Agarwal R, Kaushal M, Aggarwal R, Paul VK, Deorari AK. Early neonatal hyperbilirubinemia using first day serum bilirubin level. Indian Pediatr. 2002;39:724-30.
23. Park K. Man and Medicine: Towards Health for All: In K. Park (Eds.); Textbook of Preventive and Social Medicine. Jabalpur; Banarsidas Bhanot; 2005;18:1-11.
24. Shally Awasthi and Hasibur Rehman. Early Prediction of Neonatal Hyperbilirubinemia. Indian J Pediatr 1998;65: 131-139
25. Zakia nahar MD. Shahidukkah , Abdul Mannan, Sanjoy Kumar Dey, Ujjal Mitra, SM Selimuzzaman: the value of umbilical cord blood bilirubin measurement in predicting the development of significant hyperbilirubinemia in healthy newborn. Bangladesh J Child Health 2009; Vol 33(2):50-54